CURRENT OPINION



Modern Biomarkers for Autism Spectrum Disorder: Future Directions

Amanda R. Jensen¹ · Alison L. Lane¹ · Brianna A. Werner¹ · Sallie E. McLees¹ · Tessa S. Fletcher^{1,2} · Richard E. Frye¹

Accepted: 16 May 2022 / Published online: 27 June 2022 $\ensuremath{\textcircled{O}}$ The Author(s) 2022

Abstract

Autism spectrum disorder is an increasingly prevalent neurodevelopmental disorder in the world today, with an estimated 2% of the population being affected in the USA. A major complicating factor in diagnosing, treating, and understanding autism spectrum disorder is that defining the disorder is solely based on the observation of behavior. Thus, recent research has focused on identifying specific biological abnormalities in autism spectrum disorder that can provide clues to diagnosis and treatment. Biomarkers are an objective way to identify and measure biological abnormalities for diagnostic purposes as well as to measure changes resulting from treatment. This current opinion paper discusses the state of research of various biomarkers currently in development for autism spectrum disorder. The types of biomarkers identified include prenatal history, genetics, neurological including neuroimaging, neurophysiologic, and visual attention, metabolic including abnormalities in mitochondrial, folate, trans-methylation, and trans-sulfuration pathways, immune including autoantibodies and cytokine dysregulation, autonomic nervous system, and nutritional. Many of these biomarkers have promising preliminary evidence for prenatal and post-natal pre-symptomatic risk assessment, confirmation of diagnosis, subtyping, and treatment response. However, most biomarkers have not undergone validation studies and most studies do not investigate biomarkers with clinically relevant comparison groups. Although the field of biomarker research in autism spectrum disorder is promising, it appears that it is currently in the early stages of development.

1 Autism Spectrum Disorder: A Disorder in Need of Objective Quantitative Biomarkers

Autism spectrum disorder (ASD) is quickly growing to be one of the most significant neurodevelopmental disorders of our time. With an estimated 2% of US children affected by ASD [1], it is growing increasingly important to identify, diagnose, treat, and understand this disorder. Many factors make ASD difficult to understand and manage; chief among them is the fact that the diagnosis of ASD is solely based on behavioral observation. While there are many biological, physiological, and medical abnormalities associated with ASD, there is yet to be any way to utilize these abnormalities to develop an objective generalizable measurement to assist with the diagnosis or management of ASD. This lack of understanding of the bio-physiological processes involved in ASD not only limits diagnostic accuracy but also impacts treatment planning. The most common treatment for ASD is behavioral and educational interventions, such as applied behavior analysis and speech and/or occupational therapy. While these treatments can be beneficial, they require a large commitment of time and energy on the part of the patient and the family. Understanding the biophysiological abnormalities of ASD can help in the development of pharmacological or other therapeutics that may augment behavior and educational therapies to accelerate habilitation. Thus, an objective validated bio-physiological measure that could represent pathophysiological processes underlying ASD may have utility for both diagnosis and monitoring treatment progress.

Biomarkers are objective measures of bio-physiological abnormalities that can have many different applications such as disease diagnosis, classification of disease severity, indication of prognosis, and measuring response to therapy [2]. While there is much research centered around biomarker development in ASD, none of these biomarkers is yet validated [3]. The heterogeneity of ASD proves to complicate biomarker development [4]. This article reviews and discusses the emerging research of various potential

Richard E. Frye rfrye@phoenixchildrens.com

¹ Section on Neurodevelopmental Disorders, Barrow Neurological Institute at Phoenix Children's Hospital, 1919 E Thomas Rd, Phoenix, AZ 85016, USA

² Department of Child Health, University of Arizona College of Medicine-Phoenix, Phoenix, AZ, USA

Key Points

Autism spectrum disorder is a prevalent neurodevelopmental disorder worldwide with an estimated prevalence of about 2% of children in the USA. However, there is no objectively proven biological measurement to provide an indication of autism spectrum disorder risk or diagnosis or to indicate optimal treatment.

Preliminary findings on genetic, metabolic, immune, and neuroimaging diagnostic biomarkers show promise, especially in conjunction with additional measurements such as behavioral assessments. However, large validation trials and the use of appropriate control populations are lacking.

Although the field of biomarker research in autism spectrum disorder is promising, it appears it is currently in the early stages of development. Biomarkers to stratify autism spectrum disorder risk during the prenatal and postnatal pre-symptomatic period may be particularly helpful for starting interventions early when they might be most effective, and biomarkers to predict treatment response may expedite habilitation for those already diagnosed.

biomarkers that could advance the understanding and management of ASD. We discuss the current standing of each biomarker, limitations of the research, and pathways for future development.

2 Current Biomarkers

Autism spectrum disorder is a complicated disorder that spans many disciplines with the understanding of the disorder evolving over many decades. Before 1986, ASD research concentrated on mostly genetics and neuroimaging research with research into mitochondrial and immune disorders starting to increase after that time, followed by an increase in research into oxidative stress and toxicants in the 1990s [5]. As research has progressed, biomarkers for specific biological processes have been developed and translated from the laboratory into clinical research trials [3]. Given the diversity of the disciplines that are involved in evaluating children with ASD, a wide variety of biomarkers have been developed, including those associated with medical history and nutrition to those involved with more molecular processes such as microRNA (miRNA). This article reviews the more promising biomarkers, selected by the quality and replicability of the research that supports them. The authors selected original studies and systematic reviews for each potential biomarker. The team then discussed whether there was sufficient evidence and whether the published studies were of significant quality to be included in this current opinion paper. Biomarkers with little evidence or those supported by poor-quality studies were not included in this current opinion paper.

Table 1 summarizes the biomarkers discussed as well as their strengths and weaknesses and whether validation studies exist. Biomarkers are also depicted by their various types [3]: risk biomarkers can provide an estimate of disease risk but cannot necessarily be used for diagnosis; diagnostic biomarkers are aimed at making or confirming a diagnosis; subgroup biomarkers can indicate whether an individual already diagnosed may belong to a subset of diseased individuals with certain characteristics; and treatment biomarkers may indicate whether an already diagnosed individual may respond to a specific treatment. In addition, biomarkers may be used during different developmental periods, including prenatal, post-natal, pre-diagnosis, or post-diagnosis. Below, each biomarker category is discussed with evidence for the biomarker discussed.

2.1 Prenatal History

In a systematic review and meta-analysis involving over 40,000 ASD cases [6], maternal infection during pregnancy was linked to an increased ASD risk, especially in cases that required hospitalization for infection. One large prospective cohort study [7] in Norway found that the ASD risk increased significantly with three or more fevers after 12 weeks gestation. Multiple other retrospective case-controlled studies [8, 9] and individual cohort studies [10, 11] have supported these findings and this finding is consistent with a well-developed animal model of ASD known as the maternal immune activation model [12]. Maternal metabolic conditions have also been linked to an increased ASD risk [13, 14], as seen in a retrospective case-controlled study [15] in California that found children with ASD of women with metabolic conditions, such as obesity and diabetes mellitus, had poorer expressive language. A meta-analysis involving over 37,000 ASD cases linked gestational diabetes to an increased ASD risk among other factors [16]. While these risk factors currently do not provide diagnostic use or prediction of therapeutic response, they may be used in the future to identify children at a high risk for developing ASD.

2.2 Genetic

Because of the apparent high heritability of ASD, research focusing on its underlying genetics has long persisted [17]. Early research concentrated on structural DNA alterations while more modern studies have included gene expression and epigenetics. Clearly, understanding these underlying

Biomarker	Type (potential)	Period	Strength	Weakness	Valida- tion studies
Prenatal history Gestational infections	Risk	Prenatal	Can obtain from medical history	Not clear If specific to specific infection or if the effects of infection treatments (eg, antibiotics) have an effect. No clear treatment conse-	No
Obesity/diabetes Genetics	Risk	Prenatal	Can obtain from medical history	year treatment consequences other than standard of care.	No
Structural DNA alterations	Risk subgroup Treatment	All periods	High-throughput comprehensive genetic analysis is becoming clinically routine. Particularly helpful in difficult, refractory cases to provide prognostic information	Low yield for any single disorder and phenotype can be variable. Variants of unknown significance are common results leading to ambiguous information which can be hard to reconcile. Experi- enced geneticist is needed to interpret complex genetic information	No
Single nucleotide polymorphism	Risk Treatment	Postnatal	Single nucleotide polymorphism is generally available even on the consumer level	Lack of rigorous scientific research on risk association and treatment implications leads to dubious treatment recommendations based on results, especially at the consumer level	No
mRNA/miRNA	Diagnostic Subgroup	Post-diagnosis	Noninvasive. Some correlation with neurodevel- opment outcomes	Variable performance in validation studies. Different RNA subtypes used in various studies. Utility unclear if diagnosis already estab- lished	Yes
Methylation	Risk Diagnostic	All periods	Biologically important. Important epigenetic fac- tor than may explain inheritance and environ- mental exposures	Very variable results regarding direction of methylation abnormalities and specific gene target of methylations changes. Very complicated field given both inherited and environmental factors are at play	No
Neurological			·		
Morphology and diffusion tensor imaging	Risk Diagnostic	Pre-diagnosis Post-diagnosis	Consistent abnormalities in brain growth appear prior to diagnosis and persist into the diagnostic period. Non-invasive	Large prospective studies are needed to validate findings. MRI may require specialized centers. Limited to infants and children that can tolerate MRI scanner	No
N170	Diagnostic	Post-Diagnosis	Non-invasive. No Sedation Needed	Requires cooperation and attention to view stimuli. Utility unclear if diagnosis already established. Large prospective studies are needed to validate findings	No
MEG	Diagnostic Treatment	Post-diagnosis	Non-invasive. No Sedation Needed. Possible association with GABA signaling leading to possible prediction of treatment response	Variable protocols limit conclusions from current research. Utility unclear if diagnosis already established. Large prospective studies are needed to validate findings	No
Resting state MRI	Risk Diagnostic	Pre-diagnosis Post-diagnosis	Non-invasive	Variable protocols limit conclusions from current research. Require specialized centers. Limited to infants and children that can tolerate MRI scanner	No
Visual attention	Risk Diagnostic	Pre-diagnosis Post-diagnosis	Non-invasive	Require specialized centers. Requires cooperation and attention to view stimuli	No
Metabolic					
Transmethylation transsulphuration	Risk Diagnostic Treatment	All Periods	Only requires blood test. Possible prenatal predictive. Biologically important and possibly relevant to treatment	Requires specialized centers and equipment and not widely available clinically. May correlation with symptoms. Large prospective stud- ies are needed to validate findings	Yes

Biomarker	Type (potential)	Period	Strength	Weakness	Valida- tion studies
Mitochondrial	Subgroup Treatment	Post-diagnosis	Various biomarkers, some non-invasive. Possibly biologically important and relevant to treatment	Variable techniques and biomarkers limit conclusions from current research. Large prospective studies using a validated biomarker are needed to validate findings	No
Immune					
Maternal autism related (MAR)	Diagnostic	Prenatal	Possible prenatal prediction of ASD diagnosis with a high sensitivity for the MAR subgroup	Large prospective studies are needed to validate findings. Treatment implications unclear at this time	No
Brain autoantibodies	Treatment	Post-diagnosis	Possibly biologically important and relevant to treatment	Variable techniques and biomarkers limit conclusions from current research. Large prospective studies using a validated biomarker are needed to validate findings	No
Folate receptor alpha autoantibody	Risk Subgroup Treatment	All periods	Possible prenatal prediction of ASD diagnosis given that found also in parents May represent an ASD subgroup that is treatment responsive	May indicate distinct at-risk subgroup but lack sensitivity for ASD diagnosis. Transgenerational features require significant more investigation. Large prospective studies are needed to validate findings	No
Cytokines	Risk Subgroup Treatment	All periods	Elevated cytokines in pregnancy and neonatal period interesting as possible risk predictor. May indicate subgroup that requires specific treatment	Variable techniques and biomarkers limit conclusions from current research. Large prospective studies using a validated biomarker are needed to validate findings	No
Autonomic					
Heart rate variability	Subgroup Treatment	Post-Diagnosis	May represent an ASD subgroup that is treatment responsive	Variable techniques and biomarkers limit conclusions from current research. Large prospective studies using a validated biomarker are needed to validate findings	No
Pupillometry	Diagnostic	Post-Diagnosis	Non-invasive	Variable techniques and biomarkers limit conclusions from current research	No
Zinc	Risk Subgroup	Prenatal Post-diagnosis	Abnormal zinc level in pregnancy period is an interesting possible risk predictor with treatment implications. May indicate subgroup that requires specific treatment	Deciduous teeth typically not available until after diagnosis so fetal zinc measurements would need to be developed. Treatment implications unclear	No
Vitamin D	Risk Treatment	Prenatal Post-diagnosis	Easily obtained prenatally with routine laborato- ries and treatment implications straightforward. Biologically relevant	Target optimum vitamin D level widely debated. Large prospective studies are needed to validate findings	No
Folate	Risk	Pre-diagnosis	Easily obtained with routine laboratories	The complicated nature of various folate species is not captured in routine total folate levels and the interaction with other folate pathway abnormalities makes routine levels hard to interpret on their own	No

molecular abnormalities can have positive impacts for developing endophenotypes and response to therapies [18].

2.2.1 Structural DNA Alterations

Research into the structural DNA changes that are associated with ASD include well-known chromosomal alterations such as Down syndrome, copy number variations such as 15q11.2 microdeletion, single-gene disorders such as Phelan McDermid Syndrome, and trinucleotide repeat disorders such as Fragile X syndromes [19]. With the age of routine, clinically available, whole exome and genome sequencing upon us, the promise to identify these disorders is greeted by much enthusiasm. However, despite the high apparent heritability, empirical studies suggest that structural genetic defects account for a minority of ASD cases with empirical studies finding that a genetic disorder can be found in only about 16% of children with ASD using both a chromosomal microarray and whole exome sequencing [20]. In addition, when a variant is found, it is usually de novo rather than inherited [21-24]. In fact, there is insufficient evidence for a ASD-specific gene or a particular genetic variant with a large effect [25]. Furthermore, only a portion of individuals with prevalent genetic abnormalities such as Down or Fragile X syndrome are co-diagnosed with ASD [3], demonstrating the lack of specificity of a genetic diagnosis. The lack of good-quality outcome studies for using a structural genetic analysis in the clinical ASD field is emphasized by the need for quantitative outcome measures [26]. Although clinical improvements in management are possible with this information, this usually requires an experienced geneticist to comprehensively interpret complex genetic information [27].

Other studies have focused on common variations in the genome known as single nucleotide polymorphisms (SNPs). Attempts to develop a diagnostic classifier using large numbers (237) of SNPs have resulted in low accuracy with a range of 56–86%, potentially because of the common variation of SNPs with other factors such as ethnicity [28].

Rather than concentrating on an absolute diagnosis, other studies have identified SNPs that confer an increased risk of being diagnosed with ASD. One study of northeast Chinese Han found that synergistic interactions between SNPs on the *SHANK2* gene, a gene important for synaptogenesis and glutamate neurotransmission, increased the risk for ASD [29]. Of particular interest is the association between SNPs in folate-one carbon metabolism and ASD risk. Single nucleotide polymorphisms in *MTHFR* [30], *RFC* [31], and *MTR* [32] alone and in combination with other folate genes [33] have been associated with an increased ASD risk [31, 32]. Although still preliminary, these studies demonstrate the complicated nature of potential polygenic influences on ASD risk. Furthermore, as these genes can have downstream effects on inherited factors such as methylation, the transgenerational genomic effects need to be considered, as exemplified by a study that demonstrated that ASD risk was associated with *RFC* SNPs in the mother but not the child [34].

Other lines of research have examined the association between SNPs and ASD symptoms. Single nucleotide polymorphisms on the *OXTR* gene have been correlated with aggression, social function, and irritability [32–34]. While SNPs on the *CD38* gene, which have been translated from an animal model of ASD, have been linked to low *CD38* expression [35] and a lack of emotions [36]. Last, pharmacogenomics has promise for guiding drug therapies in ASD [37] but outcome studies provide only modest enthusiasm for predicting severe drug–drug interactions in ASD [38].

2.2.2 Messenger RNA/microRNA

Rather than focusing on structural changes in genes, other studies have focused on gene expression to better understand the molecular physiological state of the cell. While studies have examined traditional messenger RNA (mRNA) expression, more recent studies have focused on miRNA, which are important cellular regulators, and have identified high classification accuracies and correlations with neurodevelopmental measures in small cohorts [39]. Larger studies adding piwi-interacting RNA, non-coding RNA, ribosomal RNA, and oral microbial RNA appear to be the most promising. Initial studies of a large cohort established a 79% and 85% accuracy in training (n = 372) and validation (n = 84)cohorts, respectively [40]. A larger (n = 443) multicenter study that included individuals with developmental delays using salivary miRNA found much more modest results with 67% and 66% accuracy in training and validation cohorts, respectively [41]. Yet, a more recent, larger (n = 898) multicenter study examining salivary miRNAs, small nucleolar RNAs, piwi-interacting RNA, and microbial RNAs highlighted that those with ASD and gastrointestinal disorders demonstrated unique RNA profiles but did not examine diagnostic accuracy [42].

2.2.3 Methylation

Methylation patterns measured in the genome of various tissues have been investigated as another possible diagnostic tool identifying ASD. Although differences in global methylation have been inconsistent across studies in ASD, differentially methylated CpG islands have been identified that do show significant relationships [43]. Studies have suggested that differential methylated genes are enriched for ASD-associated genes across five different tissue types showing consistency across maternal and fetal blood [44]. Methylation in biological specimens from parents have been of interest to predict the likelihood of a child being born with ASD. Distinct patterns of methylation in sperm from fathers of children with ASD have been found while global DNA hypomethylation has been found in mothers of children with ASD [34, 45]. Other studies suggest pre-conceptional multivitamins could affect cord blood and fetal DNA methylation [46, 47]. Methylation as a biomarker shows a promising future for diagnostic testing in ASD. However, the complex nature of methylation and interaction with factors such as prenatal vitamin intake and other environmental exposures, such as prenatal tobacco exposure [48], makes this a very complicated area of research.

2.3 Neurological

Given that behavior and development are centered in the brain, several approaches have used neurological testing methods to identify abnormalities associated with ASD. Some of these studies, such as magnetic resonance imaging, have been limited in scope and application because of the need for cooperation of the participant, whereas more recent approaches such as examining natural visual attention may have some more promising wide-scale applications.

2.3.1 Structural Neuroimaging

After many years, consistent morphological magnetic resonance imaging findings have converged on the overgrowth of cortical [49–51] and subcortical regions, namely the amygdala [52–54], during early infancy before ASD is diagnosable. Studies have also identified enlargement of the extra-axial fluid compartment during the postnatal presymptomatic period, which correlates with severity of the eventual ASD diagnosis [55, 56].

Studies using diffusion tensor imaging, which examines the structural development of the white matter pathways, suggest that the development trajectory of white matter organization is different between 6 and 24 months of life, typically before ASD can be diagnosed, and is associated with later severity of ASD symptoms [57, 58], with older children eventually showing brain asymmetries in white matter development of key language pathways [59] and a widespread reduction in the metrics of white matter organization [60]. Structure neuroimaging studies have not looked at the diagnostic accuracy of these evaluations, but clear differences found in early brain development at a time before ASD can be diagnosed clinically make these promising avenues to pursue.

2.3.2 Neurophysiology

Evoked response studies have identified N170 latency as a possible promising biomarker for ASD. N170 latency to facial, but not object, stimuli may be prolonged in ASD [61] with latency prolongation becoming more severe with age [62]. Other studies suggest this abnormality may be related to social skills and facial memory when comparing upright versus inverted stimuli [63] and may be specific to attending to the eyes as opposed to the nose or mouth [64]. Promising results from the Autism Biomarker Consortium for Clinical Trials (ABC-CT), a large multi-site study led by the Yale Child Study Center has leading to the acceptance of N170 latency to upright human faces as an identifier of biological subgroups of ASD into the US Food and Drug Administration's Biomarker Qualification Program [65].

Magnetoencephalography studies have used a variety of techniques and neurophysiological measurements. A recent meta-analysis found evidence for prolonged M50 and M100 latencies to pure tone stimuli [66] and a recent study suggested that the M50 latencies might be used to predict treatment response to GABA-B agonists [67]. Others have studied gamma-band coherence with various auditory stimuli [68–70].

Resting state function magnetic resonance imaging has found patterns of local hyperconnectivity associated with global hypoconnectivity [71, 72]. Studies in infants prior to diagnosis have identified connectivity pathways associated with initiation of joint attention [73, 74], severity of ritualistic behavior following diagnosis [74, 75], and progressive abnormal lateralization of language networks, which starts in infancy (prior to diagnosis) and worsens with age [74, 76]. Neurophysiological studies appear intriguing and provide some insight into biological processes, but, at this point, are based on relatively small sample sizes and have not been examined in studies that allow diagnostic accuracy to be ascertained.

2.3.3 Visual Attention

Children who are later diagnosed with ASD show reduced attention to social stimuli [77] and eyes [78, 79] after 6 months of age. Studies looking at social behavior and visual attention at a young age (under 1 year) are consistent in their findings of an early decline in social behavior and reduced preference for biological motion [80], but rely on tasks meant for very young infants and lack a long-term follow-up. While visual attention shows promise as an early biomarker for ASD because of its potential for widespread application in the clinical setting, randomized trials of young children are needed to validate the biomarker.

2.4 Metabolic

2.4.1 Trans-Methylation/Trans-Sulfuration Pathways

Prior research has shown that individuals with ASD have biomarkers of oxidative stress. One of the main

manifestations is a decrease in reduced glutathione, the body's major antioxidant, along with an increase in oxidized glutathione in plasma [81-83], brain [84], and cell lines [85]. These are accompanied by other biomarkers of abnormal trans-sulfuration [86] and oxidative damage such as 3-nitrotyrosine, 3-chlorotyrosine, and 8-oxo-deoxyguanosine [87, 88]. Trans-methylation metabolism in ASD has consistently been demonstrated to be abnormal with a decrease in S-adenosylmethionine, an increase in S-adenosylhomocysteine, and a reduced S-adenosylmethionine/Sadenosylhomocysteine ratio [86, 89]. Abnormalities in transmethylation and trans-sulfuration pathways are so pervasive that they have been investigated as diagnostic markers for ASD. One study using the Fisher Discriminant Analysis found that these biomarkers could discriminate between ASD and typically developing individuals with a 97% accuracy with a follow-up study showing up to a 96% accuracy for the training dataset and 88-95% accuracy for the validation dataset [90, 91]. Functional variations of trans-sulfation deficits have been developed but remain rather preliminary [92].

Interestingly, maternal abnormalities in trans-methylation, including plasma homocysteine, adenosine, and S-adenosylmethionine are found in mothers who have offspring who developed ASD. Additionally, mothers who were high risk versus low risk for having a child with ASD could be determined with 90% accuracy using both trans-sulfuration and trans-methylation metabolites collected during the third trimester [34, 93]. Interestingly, glutathione was found to be a potential marker of a positive clinical response to treatment with methylcobalamin in a recent systematic review and meta-analysis [94].

2.4.2 Mitochondrial Metabolism

Biomarkers of mitochondrial dysfunction are prevalent in children with ASD. A systematic review examining 220 studies on various biomarkers [95] and a meta-analysis of high-quality studies [96] both found evidence for biomarkers of mitochondrial dysfunction associated with ASD with a prevalence range from 8 to 31% depending on the biomarker. A prospective controlled study measuring respiratory chain activity in buccal tissue found that a novel biomarker of the complex I-IV activity ratio was abnormally increased in 64% of patients with ASD, particularly the patients with more severe ASD [97], while a retrospective study of 76 children with ASD using the same technique found mitochondrial enzyme activity abnormalities in 62% of patients [98]. In a small cohort of ten patients and ten matched controls, respiratory chain abnormalities were found in 80% of lymphocytes [99]. Lymphoblastoid cell line models of mitochondrial dysfunction have consistently demonstrated that about one-third have elevated mitochondrial respiratory rates, a unique type of mitochondrial dysfunction associated with ASD [85, 100, 101], and elevated respiratory rates in peripheral blood mononuclear cells has been linked to the neurodevelopmental regression subtype of ASD [102–104]. The complex I–IV activity ratio obtained from the buccal swab technique has been used to select individuals with mitochondrial dysfunction who responded to a mitochondrial cocktail [105] and enzyme activity measured using the buccal swab technique has demonstrated the response of mitochondrial activity to specific supplement treatments [106].

Thus, estimates of the prevalence of mitochondrial dysfunction vary widely, particularly because of the various biomarkers used to measure mitochondrial function. Clearly, a subset of children with ASD manifest mitochondrial dysfunction but the non-standardization of measurements and the non-specific nature of biomarker of mitochondrial dysfunction have slowed progress in this area of ASD research.

2.5 Immune

2.5.1 Maternal Fetal Brain-Directed Autoantibodies

Over the last two decades, four case-control studies, two very large (≥ 200), have documented that 7–12% of children with ASD may be associated with the maternal autoantibody-related subtype of ASD [107]. These maternal autoantibodies are directed to the fetal brain, thus identifying them has the potential to intervene during pregnancy and prevent ASD from developing.

2.5.2 Brain-Directed Autoantibodies

A recent review outlined at least 25 both small-sized and medium-sized case-control studies identifying specific and non-specific brain-directed autoantibodies [108], while a recent systematic review outlined both case reports and case series in which brain autoantibodies predicted response to intravenous immunoglobulin [109]. A medium-sized open-label, prospective, baseline-controlled cohort study found that the anti-dopamine D2L receptor and anti-tubulin autoantibodies predicted treatment response to intravenous immunoglobulin [110].

2.5.3 Folate Receptor-Alpha Autoantibody

A recent meta-analysis found an increased prevalence of the folate receptor-alpha autoantibody (FRAA) in children with ASD (71%) as compared with parents of children with ASD (45%), typically developing children without ASD siblings (15%), and children with developmental delays without ASD (5%) but not typically developing siblings of children with ASD (61%) [111], suggesting that the FRAA might be a

strong heritable risk factor for ASD. The FRAA has been shown to predict response to leucovorin treatment in a recent double-blind placebo-controlled trial [112]. The prevalence in particular families and the utility as a treatment response predictor makes the FRAA a compelling biomarker.

2.5.4 Cytokines

Cytokine level abnormalities associated with ASD have been retrospectively investigated at different stages of development over the last decade. In a medium-sized retrospective study, women who had offspring who developed ASD were found to have mid-gestational elevations in cytokines [113], while another retrospective case-controlled study found that elevated interleukin (IL)-1ß and IL-4 in neonatal blood spots was associated with an increased ASD risk [114]. A systematic review and meta-analysis of case-control studies found that elevated IL-1β, IL-6, and IL-8 was linked to ASD in childhood [115]. A series of case-control studies have identified a subgroup of children with ASD who have dysregulated IL-1ß and IL-1ß/IL-10 ratio in peripheral blood monocytes [116], finding that these dysregulated cytokines are linked to periodic behavioral flares [117], non-IgEmediated food allergies [118], changes in miRNA expression [118, 119], and changes in mitochondrial respiration [119, 120]. While there is no single cytokine abnormality that has been validated in large prospective studies, abnormal cytokine profiles appear to be associated with ASD at multiple stages of development.

2.6 Autonomic

2.6.1 Heart Rate Variability

Two prospective cross-sectional study studies and two casecontrolled studies [121–124] found lower heart rate variability in adults and children with ASD. One study found that heart rate variability predicted treatment response to propranolol [125].

2.6.2 Pupillometry

A systematic review and meta-analysis confirm the statistically significant differences in pupillary response latency in ASD, although the study did highlight the significant heterogeneity of pupillometry study designs, outcomes, and quality [126].

2.7 Nutritional

A meta-analysis of case-control studies suggested children with ASD demonstrated differences in copper in hair and serum, and lower zinc concentrations in the blood [127]. An innovative biomarker that examined deciduous teeth to measure prenatal nutrient metals found that prenatal fetal manganese and zinc was reduced in ASD as compared with their monozygotic and dizygotic twin discordant for ASD [128], while another study suggested that fetal zinc-copper rhythmicity was predictive of ASD [129]. One study has linked these prenatal disruptions in nutritional metal exposure to long-term bioenergetics and language development [104].

Another important nutritional biomarker is vitamin D, as lower first-trimester vitamin D levels [130] and mid-gestation vitamin D deficiency [131] are associated with severity of ASD behaviors in offspring, and lifetime maternal vitamin D deficiency increases the risk of ASD in children [132]. Thus, the maternal vitamin D level may be a biomarker for ASD risk and severity.

Folate during pregnancy has a complex relationship to the development of ASD in the offspring. Folate deficiency during pregnancy increases the risk of the offspring developing ASD [133] while a folic acid supplement during pregnancy clearly decreases the risk of the offspring developing ASD [134]. However, two studies suggest that very high maternal folic acid blood concentrations at birth are associated with an increased risk of developing ASD [135, 136] but this notion has significant limitations when we consider that folate metabolism is more likely to be disrupted in ASD and that the type of folate within the supplement (oxidized vs reduced) is variable [137]. Indeed, studies are demonstrating that folic acid, a synthetic oxidized form of folate that is used for fortification and in most supplements, can inhibit folate metabolism, while reduced forms of folate such as leucovorin (folinic acid) or methyltetrahydrofolate do not have inhibitor effects on folate metabolism [137].

3 The Provisional State of Biomarker Development

Because of the multi-dimensional nature of ASD, biomarkers have been developed using a wide variety of techniques and approaches. Few biomarkers have undergone validation studies. For diagnostic biomarkers, salivary miRNA has, by far, undergone the largest studies, but still the particular type of miRNA used across studies and the accuracy of the biomarker varies widely across studies. Many other potentially diagnostic biomarkers have only undergone preliminary studies that are still in the optimization stages. For example, while transmethylation/trans-sulfuration pathway biomarkers appear promising, the studies remain small, lack developmentally delayed non-ASD controls, which may be the most relevant comparison population, and use a combination of measurements that still need to be optimized. Other biomarkers are very compelling but require large validation studies to be conducted. One that is extremely compelling is the maternal autoantibody-related ASD biomarker, which has the potential to identify a subset of children who will develop ASD prenatally. Neuroimaging and visual attention studies are particularly interesting as they have the potential to provide direct biologically relevant information about brain development, which appears to start before ASD is diagnosable clinically, leading to the possibility of intervening before symptoms start. Unfortunately, these measures do require cooperation of the infant and specialized centers. Other technologies such as untargeted metabolomics [138] and proteomics [139] are emerging but are still inconsistent in their findings.

Other biomarkers such as structural genetic investigations can provide an exact diagnosis, but diagnosis of the genetic syndrome, for most syndromes, overlaps incompletely with the diagnosis of ASD, making these more of an indicator of risk, albeit high risk in most cases, for ASD. Many other biomarkers provide an indication of risk of developing ASD with many of these biomarkers available in the prenatal period with potential treatment implications. For example, verification of prenatal factors such as obesity and diabetes as risk factors for ASD may implicate more aggressive management of these maternal disorders, although studies would need to verify such an approach because it is possible that other factors related to the underlying cause of the metabolic disturbances may be the driving factor rather than the diseases themselves. While more actionable biomarkers such as vitamin D might be more directly translatable, disruptions in folate and zinc, although very compelling regarding potentially translatable treatments, need to be examined more closely to better understand the complex physiology of their pathways. For example, different types of folate (reduced vs oxidized) have different metabolisms, thus the optimal compound needs to be verified in clinical studies. Of course, not all treatment recommendations require biomarkers. Indeed, some have compiled evidence from a variety of prenatal outcomes to demonstrate the importance of specific vitamins and minerals for a healthy pregnancy [140, 141].

Biomarkers that indicate important subgroups, particularly those that lead to beneficial treatment decisions, may be very useful as we still have a limited understanding of how to identify children who require specific treatments. For example, the FRAA may provide an indication for specific treatment with leucovorin (a special type of folate) while cytokine profiles may indicate those with specific immune abnormalities who require treatment and miRNA profiles may identify those children who require an evaluation for gastrointestinal disorders.

4 Limitations

It is clear that biomarker development for ASD is only in the early stages. Aside from the lack of large validation studies, one of the major limitations of most studies is the lack of inclusion of children with developmental delays without ASD, as this is the clinically relevant comparison population in which biomarkers may be most useful in order to guide treatment and surveillance recommendations. Many studies use non-sibling typically developing children as controls, which is not ideal as they are at the lowest risk of developing ASD. While such controls may provide a good starting point for developing biomarkers, they are not particularly clinically relevant as the question that doctors are faced with in the clinic is whether a child with some symptomatology has ASD, not whether a perfectly normal child has ASD. Because of the prevalence rate of ASD, screening biomarkers will be very difficult to use as they will provide a low positive predictive rate even with a high sensitivity and specificity [142]. Furthermore, many biomarkers are measured after the child has developed ASD, thus the use of such a biomarker is limited, especially when the comparison group is a typically developing child who is past the age of ASD diagnosis. While current controls have been suitable for preliminary biomarker research, delineating between these populations in more rigorous future studies will help elucidate the biological mechanisms of those with unspecified developmental delays and ensure better controls.

5 The Promise of Future Biomarker Research

The future of ASD biomarker research lies in translating findings from the biological basis of ASD into validated clinically useful biomarkers for risk assessment, diagnosis, prediction of clinically relevant subgroups, and treatment response. Research is already moving in this direction, as evidenced by studies that have developed biomarkers such as maternal autoantibody-related autism biomarkers, which can predict ASD prenatally, and genetic biomarkers, which may link specific syndromes to ASD. Studies of metabolic biomarkers may provide insight into treatment, while neuroimaging biomarkers may provide biologically relevant indicators of abnormal brain development.

Common themes in areas of future development include the replication of studies with more diverse populations, more randomized controlled trials with larger numbers of participants, and better-defined cohorts with gold standard diagnostic instruments. Another common theme for the future is further investigation of biomarkers in subgroupings of ASD, as ASD is a very heterogeneous condition that will most likely require optimized individualized treatment. In conclusion, the current state of biomarker research is still preliminary but promising.

Declarations

Funding This research was funded, in part, by the Brain Foundation (Pleasanton, CA), the O'Sullivan foundation (Princeton, NJ), the N

of 1 Foundation (Dallas TX), The Jonty Foundation (St Paul, MN), the Gupta Family Foundation (Atherton, CA), and the Jager Family Foundation (Chicago, IL).

Conflict of interest Richard E. Frye is funded by the National Institutes of Health, Department of Defense, and Autism Speaks and receives support from the Turnabout for Autism, the Brain Foundation, the Autism Research Institute and Zynerba Pharmaceuticals. He is on the advisory boards of Iliad Neurosciences and NeuroNeeds. The remaining authors declare that they have no conflicts of interest that are directly relevant to the contents of this article.

Ethics approval Not applicable.

Consent to participate Not applicable.

Consent for publication Not applicable.

Availability of data and material All the authors are accountable for all aspects of the work, including full data access, integrity of the data, and the accuracy of the data analysis. All authors will ensure that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Code availability Not applicable.

Author contributions All the authors contributed to the study conception and design. The first draft of the manuscript was written by all the authors. All the authors read and approved the final manuscript.

Open Access This article is licensed under a Creative Commons Attribution-NonCommercial 4.0 International License, which permits any non-commercial use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by-nc/4.0/.

References

- Maenner MJ, et al. Prevalence and characteristics of autism spectrum disorder among children aged 8 years: Autism and Developmental Disabilities Monitoring Network, 11 sites, United States, 2018. MMWR Surveill Summ. 2021;70(11):1–16.
- Biomarkers Definitions Working Group. Biomarkers and surrogate endpoints: preferred definitions and conceptual framework. Clin Pharmacol Ther. 2001;69(3):89–95.
- 3. Frye RE, et al. Emerging biomarkers in autism spectrum disorder: a systematic review. Ann Transl Med. 2019;7(23):792.
- 4. Frye RE. A personalized multidisciplinary approach to evaluating and treating autism spectrum disorder. J Pers Med. 2022;12(3):464.
- Rossignol DA, Frye RE. A review of research trends in physiological abnormalities in autism spectrum disorders: immune dysregulation, inflammation, oxidative stress, mitochondrial dysfunction and environmental toxicant exposures. Mol Psychiatry. 2012;17(4):389–401.

- Jiang HY, et al. Maternal infection during pregnancy and risk of autism spectrum disorders: a systematic review and metaanalysis. Brain Behav Immun. 2016;58:165–72.
- Hornig M, et al. Prenatal fever and autism risk. Mol Psychiatry. 2018;23(3):759–66.
- Zerbo O, et al. Maternal infection during pregnancy and autism spectrum disorders. J Autism Dev Disord. 2015;45(12):4015–25.
- Zerbo O, et al. Is maternal influenza or fever during pregnancy associated with autism or developmental delays? Results from the CHARGE (CHildhood Autism Risks from Genetics and Environment) study. J Autism Dev Disord. 2013;43(1):25–33.
- Lee BK, et al. Maternal hospitalization with infection during pregnancy and risk of autism spectrum disorders. Brain Behav Immun. 2015;44:100–5.
- Brucato M, et al. Prenatal exposure to fever is associated with autism spectrum disorder in the boston birth cohort. Autism Res. 2017;10(11):1878–90.
- Kwon HK, Choi GB, Huh JR. Maternal inflammation and its ramifications on fetal neurodevelopment. Trends Immunol. 2022;43(3):230–44.
- Connolly N, et al. Maternal metabolic risk factors for autism spectrum disorder: an analysis of electronic medical records and linked birth data. Autism Res. 2016;9(8):829–37.
- 14. Frye RE, et al. Mitochondria may mediate prenatal environmental influences in autism spectrum disorder. J Pers Med. 2021;11(3):218.
- Krakowiak P, et al. Maternal metabolic conditions and risk for autism and other neurodevelopmental disorders. Pediatrics. 2012;129(5):e1121–8.
- Wang C, et al. Prenatal, perinatal, and postnatal factors associated with autism: a meta-analysis. Medicine (Baltimore). 2017;96(18): e6696.
- Schaefer GB. Clinical genetic aspects of ASD spectrum disorders. Int J Mol Sci. 2016;17(2):180.
- Gill PS, et al. Molecular dysregulation in autism spectrum disorder. J Pers Med. 2021;11(9):848.
- Schaefer GB, et al. Clinical genetics evaluation in identifying the etiology of autism spectrum disorders: 2013 guideline revisions. Genet Med. 2013;15(5):399–407.
- Tammimies K, et al. Molecular diagnostic yield of chromosomal microarray analysis and whole-exome sequencing in children with autism spectrum disorder. JAMA. 2015;314(9):895–903.
- Hamdan FF, et al. High rate of recurrent de novo mutations in developmental and epileptic encephalopathies. Am J Hum Genet. 2017;101(5):664–85.
- 22. O'Roak BJ, et al. Recurrent de novo mutations implicate novel genes underlying simplex autism risk. Nat Commun. 2014;5:5595.
- 23. Wang L, et al. Functional relationships between recessive inherited genes and genes with de novo variants in autism spectrum disorder. Mol Autism. 2020;11(1):75.
- 24. Wang T, et al. Large-scale targeted sequencing identifies risk genes for neurodevelopmental disorders. Nat Commun. 2020;11(1):4932.
- Myers SM, et al. Insufficient evidence for "autism-specific" genes. Am J Hum Genet. 2020;106(5):587–95.
- Yusuf A, et al. Adaptation and validation of the Genetic Counseling Outcome Scale for autism spectrum disorders and related conditions. J Genet Couns. 2021;30(1):305–18.
- 27. Kreiman BL, Boles RG. State of the art of genetic testing for patients with autism: a practical guide for clinicians. Semin Pediatr Neurol. 2020;34: 100804.
- Skafidas E, et al. Predicting the diagnosis of autism spectrum disorder using gene pathway analysis. Mol Psychiatry. 2014;19(4):504–10.

- Qiu S, et al. SHANK1 polymorphisms and SNP-SNP interactions among SHANK family: a possible cue for recognition to autism spectrum disorder in infant age. Autism Res. 2019;12(3):375–83.
- Frustaci A, et al. Oxidative stress-related biomarkers in autism: systematic review and meta-analyses. Free Radic Biol Med. 2012;52(10):2128–41.
- Mahmuda NA, et al. A study of single nucleotide polymorphisms of the SLC19A1/RFC1 gene in subjects with autism spectrum disorder. Int J Mol Sci. 2016;17(5):772.
- Haghiri R, et al. Analysis of methionine synthase (rs1805087) gene polymorphism in autism patients in Northern Iran. Acta Neurobiol Exp (Wars). 2016;76(4):318–23.
- James SJ, et al. Metabolic endophenotype and related genotypes are associated with oxidative stress in children with autism. Am J Med Genet B Neuropsychiatr Genet. 2006;141B(8):947–56.
- 34. James SJ, et al. A functional polymorphism in the reduced folate carrier gene and DNA hypomethylation in mothers of children with autism. Am J Med Genet B Neuropsychiatr Genet. 2010;153B(6):1209–20.
- Lerer E, et al. Low CD38 expression in lymphoblastoid cells and haplotypes are both associated with autism in a family-based study. Autism Res. 2010;3(6):293–302.
- Krol KM, et al. Genetic variation in the oxytocin system and its link to social motivation in human infants. Psychoneuroendocrinology. 2021;131: 105290.
- 37. Gill PS, et al. Multidisciplinary consulting team for complicated cases of neurodevelopmental and neurobehavioral disorders: assessing the opportunities and challenges of integrating pharmacogenomics into a team setting. J Pers Med. 2022;12(4):599.
- Ariefdjohan M, et al. The utility of pharmacogenetic-guided psychotropic medication selection for pediatric patients: a retrospective study. Pediatr Rep. 2021;13(3):421–33.
- Hicks SD, et al. Salivary miRNA profiles identify children with autism spectrum disorder, correlate with adaptive behavior, and implicate ASD candidate genes involved in neurodevelopment. BMC Pediatr. 2016;16:52.
- 40. Hicks SD, et al. Validation of a salivary RNA test for childhood autism spectrum disorder. Front Genet. 2018;9:534.
- 41. Hicks SD, et al. Saliva MicroRNA differentiates children with autism from peers with typical and atypical development. J Am Acad Child Adolesc Psychiatry. 2020;59(2):296–308.
- 42. Beversdorf DQ, et al. Saliva RNA biomarkers of gastrointestinal dysfunction in children with autism and neurodevelopmental disorders: potential implications for precision medicine. Front Psychiatry. 2021;12: 824933.
- Vogel Ciernia A, LaSalle J. The landscape of DNA methylation amid a perfect storm of autism aetiologies. Nat Rev Neurosci. 2016;17(7):411–23.
- 44. Bakulski KM, et al. Autism-associated DNA methylation at birth from multiple tissues is enriched for autism genes in the early autism risk longitudinal investigation. Front Mol Neurosci. 2021;14: 775390.
- 45. Garrido N, et al. Sperm DNA methylation epimutation biomarker for paternal offspring autism susceptibility. Clin Epigenet. 2021;13(1):6.
- 46. Bakulski KM, et al. Prenatal multivitamin use and MTHFR genotype are associated with newborn cord blood DNA methylation. Int J Environ Res Public Health. 2020;17(24):9190.
- McKay JA, et al. Genetic and non-genetic influences during pregnancy on infant global and site specific DNA methylation: role for folate gene variants and vitamin B12. PLoS ONE. 2012;7(3): e33290.
- Hannon E, et al. Elevated polygenic burden for autism is associated with differential DNA methylation at birth. Genome Med. 2018;10(1):19.

- 49. Dierker DL, et al. Analysis of cortical shape in children with simplex autism. Cereb Cortex. 2015;25(4):1042–51.
- Hazlett HC, et al. Early brain development in infants at high risk for autism spectrum disorder. Nature. 2017;542(7641):348–51.
- Pote I, et al. Familial risk of autism alters subcortical and cerebellar brain anatomy in infants and predicts the emergence of repetitive behaviors in early childhood. Autism Res. 2019;12(4):614–27.
- 52. Nordahl CW, et al. Increased rate of amygdala growth in children aged 2 to 4 years with autism spectrum disorders: a longitudinal study. Arch Gen Psychiatry. 2012;69(1):53–61.
- Barnea-Goraly N, et al. A preliminary longitudinal volumetric MRI study of amygdala and hippocampal volumes in autism. Prog Neuropsychopharmacol Biol Psychiatry. 2014;48:124–8.
- Zhu Z, et al. Alterations in volumes and MRI features of amygdala in Chinese autistic preschoolers associated with social and behavioral deficits. Brain Imaging Behav. 2018;12(6):1814–21.
- Shen MD, et al. Extra-axial cerebrospinal fluid in high-risk and normal-risk children with autism aged 2–4 years: a case-control study. Lancet Psychiatry. 2018;5(11):895–904.
- Shen MD, et al. Early brain enlargement and elevated extra-axial fluid in infants who develop autism spectrum disorder. Brain. 2013;136(Pt 9):2825–35.
- Wolff JJ, et al. Differences in white matter fiber tract development present from 6 to 24 months in infants with autism. Am J Psychiatry. 2012;169(6):589–600.
- Wolff JJ, et al. Neural circuitry at age 6 months associated with later repetitive behavior and sensory responsiveness in autism. Mol Autism. 2017;8:8.
- Joseph RM, et al. Structural asymmetries of language-related gray and white matter and their relationship to language function in young children with ASD. Brain Imaging Behav. 2014;8(1):60–72.
- Walker L, et al. Diffusion tensor imaging in young children with autism: biological effects and potential confounds. Biol Psychiatry. 2012;72(12):1043–51.
- 61. Sysoeva OV, Constantino JN, Anokhin AP. Event-related potential (ERP) correlates of face processing in verbal children with autism spectrum disorders (ASD) and their first-degree relatives: a family study. Mol Autism. 2018;9:41.
- Kang E, et al. Atypicality of the N170 event-related potential in autism spectrum disorder: a meta-analysis. Biol Psychiatry Cogn Neurosci Neuroimaging. 2018;3(8):657–66.
- Webb SJ, et al. ERP responses differentiate inverted but not upright face processing in adults with ASD. Soc Cogn Affect Neurosci. 2012;7(5):578–87.
- 64. Parker TC, et al. The N170 event-related potential reflects delayed neural response to faces when visual attention is directed to the eyes in youths with ASD. Autism Res. 2021;14(7):1347–56.
- 65. Shic F, et al. The autism biomarkers consortium for clinical trials: evaluation of a battery of candidate eye-tracking biomarkers for use in autism clinical trials. Mol Autism. 2022;13(1):15.
- 66. Williams ZJ, et al. Cortical auditory processing of simple stimuli is altered in autism: a meta-analysis of auditory evoked responses. Biol Psychiatry Cogn Neurosci Neuroimaging. 2021;6(8):767–81.
- Roberts TPL, Kuschner ES, Edgar JC. Biomarkers for autism spectrum disorder: opportunities for magnetoencephalography (MEG). J Neurodev Disord. 2021;13(1):34.
- 68. Seymour RA, et al. Reduced auditory steady state responses in autism spectrum disorder. Mol Autism. 2020;11(1):56.
- 69. Roberts TPL, et al. Magnetoencephalography studies of the envelope following response during amplitude-modulated sweeps: diminished phase synchrony in autism spectrum disorder. Front Hum Neurosci. 2021;15: 787229.

- Port RG, et al. Maturation of auditory neural processes in autism spectrum disorder: a longitudinal MEG study. Neuroimage Clin. 2016;11:566–77.
- Uddin LQ, et al. Salience network-based classification and prediction of symptom severity in children with autism. JAMA Psychiat. 2013;70(8):869–79.
- Li D, Karnath HO, Xu Z. Candidate biomarkers in children with autism spectrum disorder: a review of MRI studies. Neurosci Bull. 2017;33(2):219–37.
- Eggebrecht AT, et al. Joint attention and brain functional connectivity in infants and toddlers. Cereb Cortex. 2017;27(3):1709–20.
- 74. Hiremath CS, et al. Emerging behavioral and neuroimaging biomarkers for early and accurate characterization of autism spectrum disorders: a systematic review. Transl Psychiatry. 2021;11(1):42.
- McKinnon CJ, et al. Restricted and repetitive behavior and brain functional connectivity in infants at risk for developing autism spectrum disorder. Biol Psychiatry Cogn Neurosci Neuroimaging. 2019;4(1):50–61.
- Eyler LT, Pierce K, Courchesne E. A failure of left temporal cortex to specialize for language is an early emerging and fundamental property of autism. Brain. 2012;135(Pt 3):949–60.
- Bradshaw J, et al. Development of attention from birth to 5 months in infants at risk for autism spectrum disorder. Dev Psychopathol. 2020;32(2):491–501.
- Shultz S, Klin A, Jones W. Neonatal transitions in social behavior and their implications for autism. Trends Cogn Sci. 2018;22(5):452–69.
- Jones W, Klin A. Attention to eyes is present but in decline in 2-6-month-old infants later diagnosed with autism. Nature. 2013;504(7480):427–31.
- Mason L, et al. Preference for biological motion is reduced in ASD: implications for clinical trials and the search for biomarkers. Mol Autism. 2021;12(1):74.
- Chen L, et al. Oxidative stress marker aberrations in children with autism spectrum disorder: a systematic review and meta-analysis of 87 studies (N = 9109). Transl Psychiatry. 2021;11(1):15.
- Main PA, et al. The potential role of the antioxidant and detoxification properties of glutathione in autism spectrum disorders: a systematic review and meta-analysis. Nutr Metab (Lond). 2012;9:35.
- Ghanizadeh A, et al. Glutathione-related factors and oxidative stress in autism, a review. Curr Med Chem. 2012;19(23):4000–5.
- Rose S, et al. Evidence of oxidative damage and inflammation associated with low glutathione redox status in the autism brain. Transl Psychiatry. 2012;2(7): e134.
- Rose S, et al. Oxidative stress induces mitochondrial dysfunction in a subset of autism lymphoblastoid cell lines in a well-matched case control cohort. PLoS ONE. 2014;9(1): e85436.
- Guo BQ, Li HB, Ding SB. Blood homocysteine levels in children with autism spectrum disorder: an updated systematic review and meta-analysis. Psychiatry Res. 2020;291: 113283.
- Frye RE, et al. Redox metabolism abnormalities in autistic children associated with mitochondrial disease. Transl Psychiatry. 2013;3(6): e273.
- Ghezzo A, et al. Oxidative stress and erythrocyte membrane alterations in children with autism: correlation with clinical features. PLoS ONE. 2013;8(6): e66418.
- Guo BQ, Ding SB, Li HB. Blood biomarker levels of methylation capacity in autism spectrum disorder: a systematic review and meta-analysis. Acta Psychiatr Scand. 2020;141(6):492–509.
- 90. Howsmon DP, et al. Classification and adaptive behavior prediction of children with autism spectrum disorder based upon multivariate data analysis of markers of oxidative stress and DNA methylation. PLoS Comput Biol. 2017;13(3): e1005385.

- Howsmon DP, et al. Multivariate techniques enable a biochemical classification of children with autism spectrum disorder versus typically-developing peers: a comparison and validation study. Bioeng Transl Med. 2018;3(2):156–65.
- Alberti A, et al. Sulphation deficit in "low-functioning" autistic children: a pilot study. Biol Psychiatry. 1999;46(3):420–4.
- Hollowood K, et al. Maternal metabolic profile predicts high or low risk of an autism pregnancy outcome. Res Autism Spectr Disord. 2018;56:72–82.
- 94. Rossignol DA, Frye RE. The effectiveness of cobalamin (B12) treatment for autism spectrum disorder: a systematic review and meta-analysis. J Pers Med. 2021;11(8):784.
- Rose S, et al. Clinical and molecular characteristics of mitochondrial dysfunction in autism spectrum disorder. Mol Diagn Ther. 2018;22(5):571–93.
- Rossignol DA, Frye RE. Mitochondrial dysfunction in autism spectrum disorders: a systematic review and meta-analysis. Mol Psychiatry. 2012;17(3):290–314.
- Goldenthal MJ, et al. Mitochondrial enzyme dysfunction in autism spectrum disorders; a novel biomarker revealed from buccal swab analysis. Biomark Med. 2015;9(10):957–65.
- Delhey L, et al. Bioenergetic variation is related to autism symptomatology. Metab Brain Dis. 2017;32(6):2021–31.
- 99. Giulivi C, et al. Mitochondrial dysfunction in autism. JAMA. 2010;304(21):2389–96.
- Bennuri SC, Rose S, Frye RE. Mitochondrial dysfunction is inducible in lymphoblastoid cell lines from children with autism and may involve the TORC1 pathway. Front Psychiatry. 2019;10:269.
- Rose S, et al. Oxidative stress induces mitochondrial dysfunction in a subset of autistic lymphoblastoid cell lines. Transl Psychiatry. 2014;4(4): e377.
- Singh K, et al. Developmental regression and mitochondrial function in children with autism. Ann Clin Transl Neurol. 2020;7(5):683–94.
- Frye RE, et al. Prenatal air pollution influences neurodevelopment and behavior in autism spectrum disorder by modulating mitochondrial physiology. Mol Psychiatry. 2021;26(5):1561–77.
- Frye RE, et al. Early life metal exposure dysregulates cellular bioenergetics in children with regressive autism spectrum disorder. Transl Psychiatry. 2020;10(1):223.
- 105. Legido A, et al. Effect of a combination of carnitine, coenzyme Q10 and alpha-lipoic acid (MitoCocktail) on mitochondrial function and neurobehavioral performance in children with autism spectrum disorder. In: 17th annual meeting of the international society for autism research; 9–12 May 2018; Rotterdam.
- Delhey LM, et al. The effect of mitochondrial supplements on mitochondrial activity in children with autism spectrum disorder. J Clin Med. 2017;6(2):18.
- 107. Edmiston E, Ashwood P, Van de Water J. Autoimmunity, autoantibodies, and autism spectrum disorder. Biol Psychiatry. 2017;81(5):383–90
- Hughes HK, et al. Immune dysfunction and autoimmunity as pathological mechanisms in autism spectrum disorders. Front Cell Neurosci. 2018;12:405.
- Rossignol DA, Frye RE. A systematic review and meta-analysis of immunoglobulin G abnormalities and the therapeutic use of intravenous limunoglobulins (IVIG) in autism spectrum disorder. J Pers Med. 2021;11(6):488.
- Connery K, et al. Intravenous immunoglobulin for the treatment of autoimmune encephalopathy in children with autism. Transl Psychiatry. 2018;8(1):148.
- 111. Rossignol DA, Frye RE. Cerebral folate deficiency, folate receptor alpha autoantibodies and leucovorin (folinic acid) treatment in autism spectrum disorders: a systematic review and metaanalysis. J Pers Med. 2021;11(11):1141.

494

- 112. Rose S, et al. Butyrate enhances mitochondrial function during oxidative stress in cell lines from boys with autism. Transl Psychiatry. 2018;8(1):42.
- 113. Jones KL, et al. Autism with intellectual disability is associated with increased levels of maternal cytokines and chemokines during gestation. Mol Psychiatry. 2017;22(2):273–9.
- Krakowiak P, et al. Neonatal cytokine profiles associated with autism spectrum disorder. Biol Psychiatry. 2017;81(5):442–51.
- Masi A, et al. Cytokine aberrations in autism spectrum disorder: a systematic review and meta-analysis. Mol Psychiatry. 2015;20(4):440–6.
- 116. Jyonouchi H, Geng L. Associations between monocyte and T cell cytokine profiles in autism spectrum disorders: effects of dysregulated innate immune responses on adaptive responses to recall antigens in a subset of ASD Children. Int J Mol Sci. 2019;20(19):4731.
- 117. Jyonouchi H, Geng L, Davidow AL. Cytokine profiles by peripheral blood monocytes are associated with changes in behavioral symptoms following immune insults in a subset of ASD subjects: an inflammatory subtype? J Neuroinflamm. 2014;11:187.
- 118. Jyonouchi H, et al. MicroRNA expression changes in association with changes in interleukin-1ss/interleukin10 ratios produced by monocytes in autism spectrum disorders: their association with neuropsychiatric symptoms and comorbid conditions (observational study). J Neuroinflamm. 2017;14(1):229.
- Jyonouchi H, et al. Serum microRNAs in ASD: association with monocyte cytokine profiles and mitochondrial respiration. Front Psychiatry. 2019;10:614.
- Jyonouchi H, et al. Variations in mitochondrial respiration differ in IL-1ss/IL-10 ratio based subgroups in autism spectrum disorders. Front Psychiatry. 2019;10:71.
- 121. Thapa R, et al. Reduced heart rate variability in adults with autism spectrum disorder. Autism Res. 2019;12(6):922–30.
- Lory C, et al. Brief report: reduced heart rate variability in children with autism spectrum disorder. J Autism Dev Disord. 2020;50(11):4183–90.
- 123. Gonzaga CN, et al. Autonomic responses to facial expression tasks in children with autism spectrum disorders: cross-section study. Res Dev Disabil. 2021;116: 104034.
- 124. Bharath R, et al. Comparison of physiological and biochemical autonomic indices in children with and without autism spectrum disorders. Medicina (Kaunas). 2019;55(7):346.
- Zamzow RM, et al. Effects of acute beta-adrenergic antagonism on verbal problem solving in autism spectrum disorder and exploration of treatment response markers. J Clin Exp Neuropsychol. 2017;39(6):596–606.
- de Vries L, et al. Autism spectrum disorder and pupillometry: a systematic review and meta-analysis. Neurosci Biobehav Rev. 2021;120:479–508.

- 127. Zhang J, et al. Trace elements in children with autism spectrum disorder: a meta-analysis based on case-control studies. J Trace Elem Med Biol. 2021;67: 126782.
- 128. Arora M, et al. Fetal and postnatal metal dysregulation in autism. Nat Commun. 2017;8:15493.
- 129. Curtin P, et al. Dynamical features in fetal and postnatal zinccopper metabolic cycles predict the emergence of autism spectrum disorder. Sci Adv. 2018;4(5):eaat1293.
- Chen J, et al. Lower maternal serum 25(OH) D in first trimester associated with higher autism risk in Chinese offspring. J Psychosom Res. 2016;89:98–101.
- 131. Vinkhuyzen AAE, et al. Gestational vitamin D deficiency and autism spectrum disorder. BJPsych Open. 2017;3(2):85–90.
- Magnusson C, et al. Maternal vitamin D deficiency and the risk of autism spectrum disorders: population-based study. BJPsych Open. 2016;2(2):170–2.
- Ornoy A, Weinstein-Fudim L, Ergaz Z. Prenatal factors associated with autism spectrum disorder (ASD). Reprod Toxicol. 2015;56:155–69.
- 134. Levine SZ, et al. Association of maternal use of folic acid and multivitamin supplements in the periods before and during pregnancy with the risk of autism spectrum disorder in offspring. JAMA Psychiat. 2018;75(2):176–84.
- 135. Raghavan R, et al. Maternal multivitamin intake, plasma folate and vitamin B12 levels and autism spectrum disorder risk in offspring. Paediatr Perinat Epidemiol. 2018;32(1):100–11.
- 136. Egorova O, et al. Maternal blood folate status during early pregnancy and occurrence of autism spectrum disorder in offspring: a study of 62 serum biomarkers. Mol Autism. 2020;11(1):7.
- Frye RE, Slattery JC, Quadros EV. Folate metabolism abnormalities in autism: potential biomarkers. Biomark Med. 2017;11(8):687–99.
- 138. Brister D, et al. Central nervous system metabolism in autism, epilepsy and developmental delays: a cerebrospinal fluid analysis. Metabolites. 2022;12(5):371.
- 139. Mesleh AG, Abdulla SA, El-Agnaf O. Paving the way toward personalized medicine: current advances and challenges in multi-OMICS approach in autism spectrum disorder for biomarkers discovery and patient stratification. J Pers Med. 2021;11(1):41.
- Adams JB, et al. Evidence-based recommendations for an optimal prenatal supplement for women in the U.S., part two: minerals. Nutrients. 2021;13(6):1849.
- 141. Adams JB, et al. The science behind the healthy child guide. Dallas: Neurological Health Foundation; 2020.
- McCarty P, Frye RE. Early detection and diagnosis of autism spectrum disorder: why is it so difficult? Semin Pediatr Neurol. 2020;35: 100831.